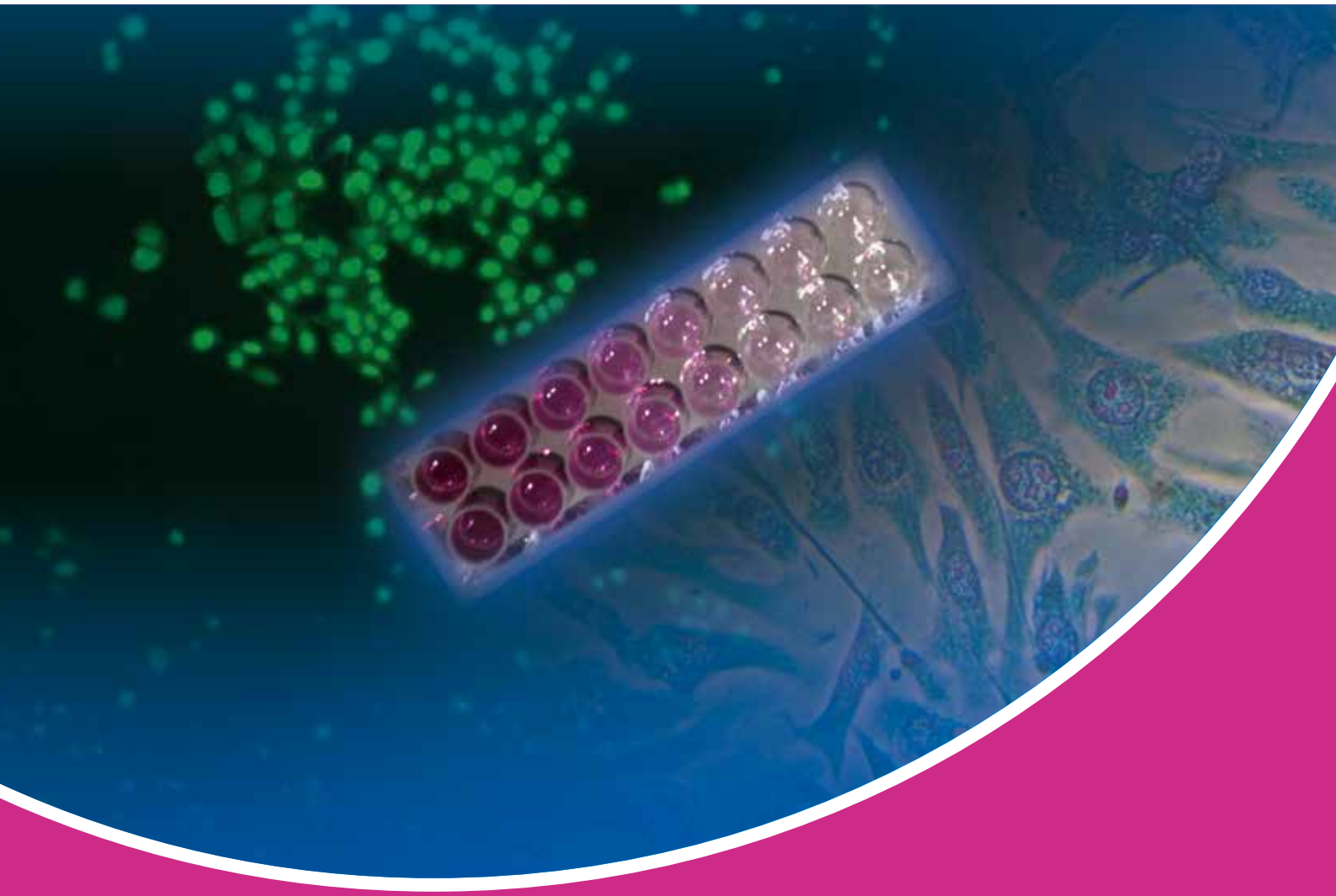
LIFE SCIENCE

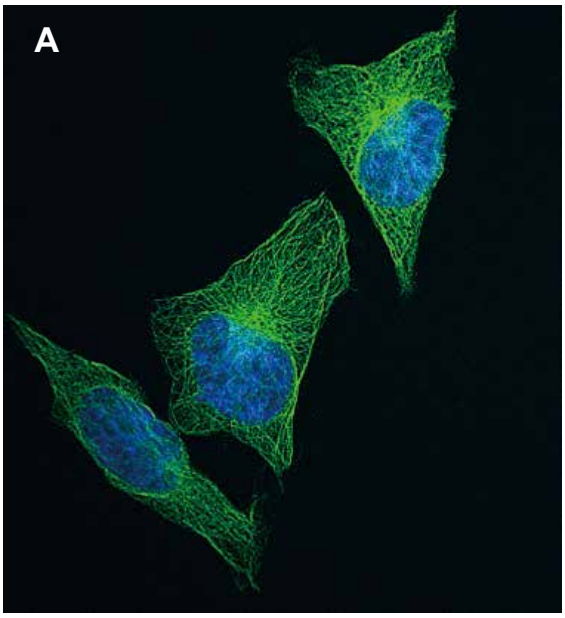
TCI

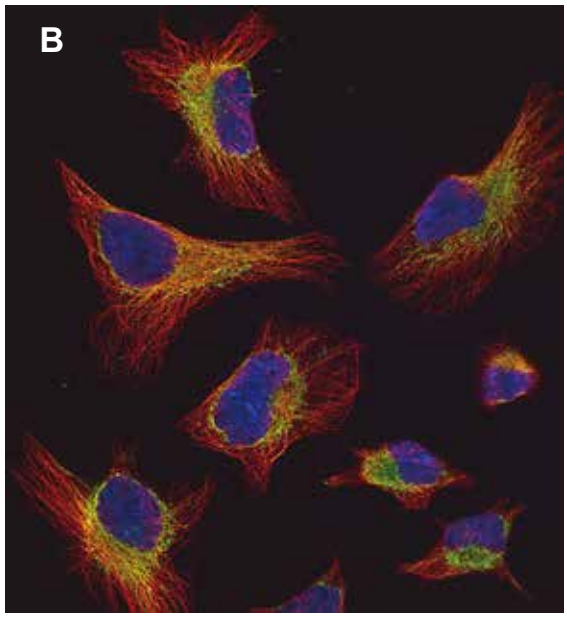
Cell Biology Reagents



Fluorescent Labeled Antibodies / Streptavidins and Fluorescent Nuclear Stains

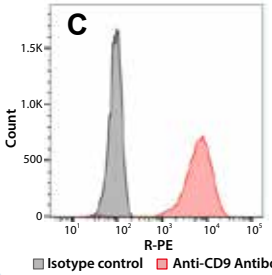
Applications





(A) The HeLa cells were incubated with properly diluted primary antibody (Mouse Anti α-Tubulin IgG) and were further incubated with Goat Anti-Mouse IgG Biotin Conjugate [G0387] and Streptavidin FITC Conjugate [S0966] (green fluorescence). And then the nuclei was stained with DAPI 2HCl [A2412] (blue fluorescence). (Laser Scanning Microscope: Olympus FLUOVIEW FV 3000)

(B) The nuclei of HeLa cells was stained with Bisbenzimide H 33258 [H1343] (blue fluorescence). α-Tubulin was stained with anti-α-tubulin antibody and Goat Anti-Mouse IgG Biotin Conjugate [G0387] and Streptavidin R-PE Conjugate [T3885] (red fluorescence). Mitochondria was stained with primary antibody and Goat Anti-Rabbit IgG FITC Conjugate [G0452] (green fluorescence)**. (Laser Scanning Microscope: Olympus FLUOVIEW FV 3000)



(C) The HeLa cells were incubated with Mouse Anti-CD9 Antibody (red line) or Mouse IgG2aκ isotype control (black line). Subsequently, both were stained with Goat Anti-Mouse IgG Biotin Conjugate [G0387] and Streptavidin R-PE Conjugate [T3885]. (Flow cytometer: Sysmex RF-500)

**Please refer to our product page for staining procedure.
R-PE/FITC-labeled anti-Mouse IgG or anti-Rabbit IgG antibodies and streptavidins can be used for fluorescence immunostaining and flow cytometry.

Goat Anti-Mouse IgG FITC Conjugate	(Green Fluorescence)	0.1mg/vial	[G0406]
Goat Anti-Mouse IgM FITC Conjugate	(Green Fluorescence)	0.1mg/vial	[G0453]
Goat Anti-Rabbit IgG FITC Conjugate	(Green Fluorescence)	0.1mg/vial	[G0452]
Streptavidin FITC Conjugate	(Green Fluorescence)	0.1mg/vial	[S0966]
Goat Anti-Mouse IgG R-PE Conjugate	(Red Fluorescence)	0.1mg/vial	[G0569]
Goat Anti-Mouse IgG_1 Fab Fragment Cyanine 3 Conjugate	(Red Fluorescence)	0.05mg/vial	[G0598]
Goat Anti-Rabbit IgG R-PE Conjugate	(Red Fluorescence)	0.1mg/vial	[G0577]
Streptavidin R-PE Conjugate	(Red Fluorescence)	0.1mg/vial	[T3885]
Goat Anti-Mouse IgG DTBTA-Eu^{3+} Conjugate	(Red Fluorescence)	0.1mg/vial	[G0505]
Goat Anti-Rabbit IgG DTBTA-Eu^{3+} Conjugate	(Red Fluorescence)	0.1mg/vial	[G0506]
Streptavidin DTBTA-Eu^{3+} Conjugate	(Red Fluorescence)	0.1mg/vial	[S0993]
DAPI 2HCl	(Blue Fluorescence)	5mg	[A2412]
DAPI 2HCl (1mg/mL in Water)	(Blue Fluorescence)	0.2mL x 5vial	[D5888]
Bisbenzimide H 33258 Hydrate	(Blue Fluorescence)	25mg	[H1343]
Bisbenzimide H 33258 (1mg/mL in Water)	(Blue Fluorescence)	0.2mL x 5vial	[B6236]

*Some products are unavailable in the Americas and China.
*The high-sensitivity detection of DTBTA-Eu^{3+} labeled probes requires time-resolved fluorometry.

Please inquire for pricing and availability of listed products to our local sales representatives.

Lipid Droplet Fluorescent Stains

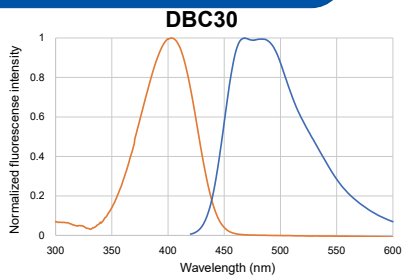
DBC30 (2mg/mL in Dimethyl Sulfoxide) [for Biochemical Research] 0.1mL [D6131]

PC6S (1mg/mL in Dimethyl Sulfoxide) [for Biochemical Research]
0.04mL / 0.2mL [P3152]

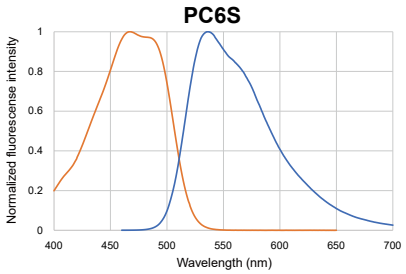
Advantages

- **Stains lipid droplets in both cells and tissues clearly and distinctly due to the high fluorescence intensity.**
- **Can be used in combination with multiple fluorescent dyes due to the high lipid droplet selectivity and intracellular retention.**

Excitation and Emission Spectra

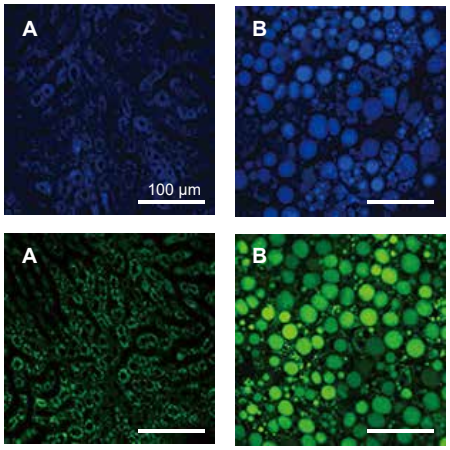


$\lambda_{ex\ max}$ = 403 nm, $\lambda_{em\ max}$ = 468 nm (in dibutyl ether)
Compatible with optical filter for DAPI.



$\lambda_{ex\ max}$ = 468 nm, $\lambda_{em\ max}$ = 535 nm (in dibutyl ether)
Compatible with optical filter for GFP or FITC.

Application : *in vivo* imaging of lipid droplets



The images of the surface of the mouse liver

Generation of fatty liver model mouse

- Feed Choline-deficient and L-Methionine-reduced ultra-high fat diet.

Lipid droplet imaging with DBC30 or PC6S

- Inject 50 nmol of DBC30 or PC6S into the tail vain of the mouse under anesthesia.
- After 30 minutes, expose the liver of the mouse and observe through confocal fluorescence microscope.

DBC30 Excitation wavelength : 405 nm
Observation wavelength : 440 - 480 nm

PC6S Excitation wavelength : 488 nm
Observation wavelength : 500 - 540 nm

A : Healthy mouse
B : Fatty liver model mouse

These figures are provided by Dr. Toshitada Yoshihara.

References T. Yoshihara *et al.*, *Anal. Chem.* **2020**, *92*, 4996. https://doi.org/10.1021/acs.analchem.9b05184
K. Purevsuren *et al.*, *J. Photochem. Photobiol. A* **2023**, *438*, 114562. https://doi.org/10.1016/j.jphotochem.2023.114562

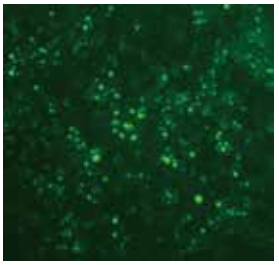
D6131 and P3152 have been commercialized under the instruction of Dr. Toshitada Yoshihara.

Pyrromethene 546

1g [D4341]

Application

1. Induce differentiation of 3T3-L1 cells into adipocytes.
2. Add 500 μL of D4341 solution prepared to 1 μM per well to stain lipid droplets.
3. After staining, fix with 4% PFA and observe.



3T3-L1 cells stained by D4341

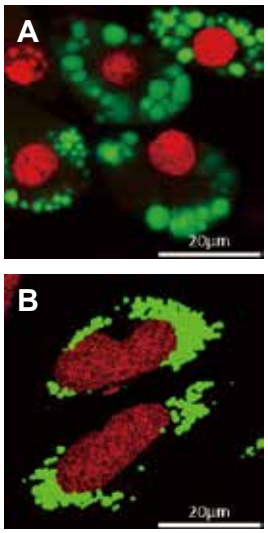
ShoyakuGreen (= TFMAQ-8Ph)

2mg [T3935]

Advantages

- **Maximum excitation wavelength at 404 nm, Maximum emission wavelength at 473 nm (in *n*-hexane)**
- **Strong and specific fluorescence imaging of lipid droplets**
- **Low cytotoxicity**

Application: Intracellular Lipid Droplet Imaging



(Observation via Leica TCS SP5)

[Cell Pre-Treatment]

(A) 3T3-L1 cells were incubated with differentiation medium in a 5% CO_2 incubator at 37°C for 10 days.

(B) HeLa cells were incubated with oleic acid in a 5% CO_2 incubator at 37°C for 48 hours.

[Preparation of ShoyakuGreen Solution]

Stock solution: ShoyakuGreen was dissolved in 100% DMSO at a concentration of 1 mM.

Working solution: The above stock solution was diluted to a final working concentration of 1 μM in culture medium.

[Fluorescence Staining of Intracellular Lipid Droplets]

1. Remove culture medium and Replace with 1 μM ShoyakuGreen working solution (Green).
2. Incubate cells in a 5% CO_2 incubator at 37°C for 30 minutes.
3. Wash – PBS x 3.
4. Fixation - 4% PFA/PB, RT, 10 minutes.
5. Wash – PBS x 3.
6. Permeabilization - 0.1% Triton X-100/PBS, RT, 15 minutes.
7. Wash – PBS x 3.
8. RNase Treatment – 100 μg/mL RNase/PBS, 37°C, 20 minutes.
9. Wash – PBS x 3.
10. PI Staining – add 5 μg/mL PI/PBS (red), 15 minutes.
11. Wash – PBS x 3.
12. Observe lipid droplets using an appropriate fluorescence microscope.

Reference Y. Fuchi, K. Hamada, S. Karasawa, *et al.*, *Sci. Rep.* **2019**, *9*, 17723. https://doi.org/10.1038/s41598-019-53882-z

T3935 has been commercialized under the instruction of Prof. Satoru Karasawa, Dr. Yasufumi Fuchi, and Dr. Koichi Hamada.

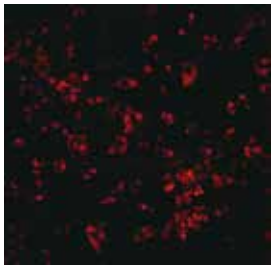
Nile Red [for Biochemical Research]

100mg / 500mg [N1272]

Nile Red is a lipophilic dye widely used to stain lipid droplets. When incorporated into intracellular lipid droplets, it emits strong fluorescence (Ex_{max}: 551 nm, Em_{max}: 631 nm). It can be used on both living and fixed cells.

Application : Lipid Droplet Staining by N1272

1. Culture 3T3-L1 cells and induce differentiation into adipocytes.
2. Remove the medium and wash twice with PBS(-).
3. Fix with 4% PFA for 10 minutes and wash twice with PBS(-).
4. Add PBS(-) containing N1272 dissolved in DMSO to a final concentration of 1 μM and incubate at 37°C for 30 minutes.
5. Remove the staining solution and wash twice with PBS(-).
6. Apply a mounting agent the cells and observe via a fluorescence microscope.



3T3-L1 cells stained by N1272

Nucleic Acid Fluorescent Stain

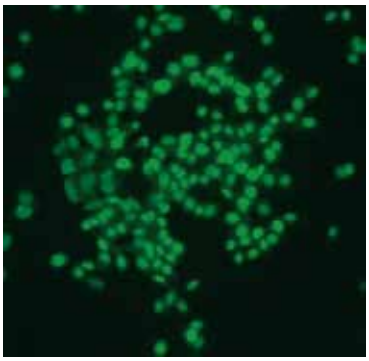
Acridine Orange Solution [for Cell Staining] 5mL [A3396]

Acridine orange is a nucleic acid staining dye that is used to identify dead cells. It intercalates with the double-stranded DNA base pairs at a ratio of 1:3 and is capable of emitting green fluorescence (Ex: 500 nm, Em: 520 nm). It also emits red fluorescence (Ex: 460 nm, Em: 650 nm) when bound to RNA or single-stranded DNA. Since acridine orange is mutagenic, this product is supplied as a solution that prevents it from splashing during weighing.

Application: The method of staining cells by A3396

1. Remove the medium from the culture plate and wash the cells twice with cold PBS(-). Remove the PBS(-).
2. Add PBS(-) and A3396 (1/50th of the volume of the added PBS(-)) and incubate for 15 minutes.
3. Remove the staining solution and wash the cells twice with PBS(-).
4. Add PBS(-) and observe the cells under a fluorescence microscope.

Please adjust the staining duration and the volume of the solution according to the cell density.
Some cells may require prior fixation; therefor optimization of the protocol according to your need is recommended.



NIH3T3 cells stained by A3396

Cell Membrane Fluorescent Stains

DiI Solution [for Cell Membrane Staining] 1mL [D6202]

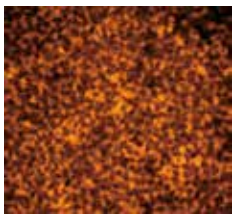
DiD Solution [for Cell Membrane Staining] 1mL [D6263]

Advantages

- **Since these products are comprised of premade solution, simply add to culture medium to stain cell membranes.**
- **Once the hydrocarbon chains are incorporated into cell membranes, DiI emits orange fluorescence (Ex_{max}: 553 nm, Em_{max}: 575 nm) and DiD emits red fluorescence (Ex_{max}: 657 nm, Em_{max}: 678 nm) strongly.**

Application: Cell Membrane Staining by D6202

1. Culture cells and remove the medium.
2. Add medium supplemented with D6202 to a final concentration of 5 µM, and incubate at 37°C for 10 minutes.
3. Remove the staining medium and wash three times with PBS(-).
4. Observe cell membranes using a fluorescence microscope.



HeLa cells stained by D6202

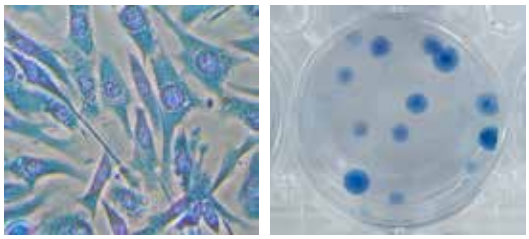
Cell Staining Dyes

Methylene Blue Solution (Methanol Solution) [for Cell Staining] 100mL [M2392]

Methylene Blue stains nuclei blue. M2392 fixes and stains cells at the same time, so you can save fixation before staining.

Application

1. Culture cells in a 6-well plate.
2. Remove medium from the plate and wash it with PBS(-) twice.
3. Remove PBS(-) from it, add 1mL of M2392 and stain cells for 15 minutes.
4. Remove M2392 from it and wash it with deionized water twice.

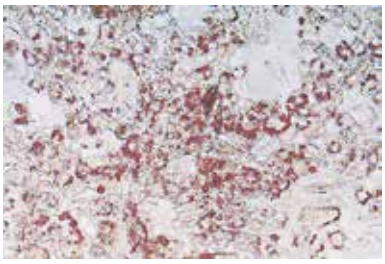


NIH/3T3 cells stained by M2392

Please adjust staining time and volume according to cells.
Because some cells need to be fixed separately, preliminary tests should be performed.

Oil Red O [for Biochemical Research] 25g [O0483]

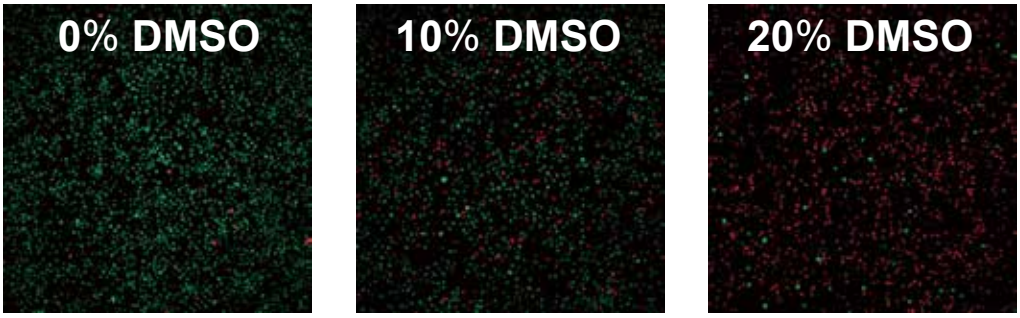
Oil red O is a diazo dye which has been widely used for staining fat cell and lipids since long ago. This lysochrome (fat-soluble dye) red dye can be used for staining of neutral triglycerides and low polar lipids. Oil Red O staining is done on fresh or frozen samples, since alcohol fixation removes lipid. Its usage is simple, and requires only washing after adding the staining solution, which makes it easy to identify lipids visually. Furthermore, accumulation of lipids can be quantified by eluting the dye using isopropanol after staining and measuring the absorbance.



10 days later of addition of media for differentiation to 3T3-L1 cell and then stained by 1mg/mL of O0483.

Live/Dead Cell Staining Kit 1kit [L0465]

L0465 (Live/Dead Cell Staining Kit) contains Calcein-AM – which stains live cells with green fluorescence through the action of intracellular esterases, and PI – which specifically stains cells with compromised cell membranes (i.e. dead cells) with red fluorescence. L0465 can help quickly, easily, and accurately distinguish between live and dead cells.



K562 cells were treated with 0-20% DMSO and stained using L0465.
At higher concentrations of DMSO, a decrease in the number of live cells (green fluorescence) is observed with a concurrent increase in the number of dead cells (red fluorescence).

Cell Proliferation/Viability Assay Reagents

ATP-Luciferase Cell Viability Assay Solution 10mL [A3519]

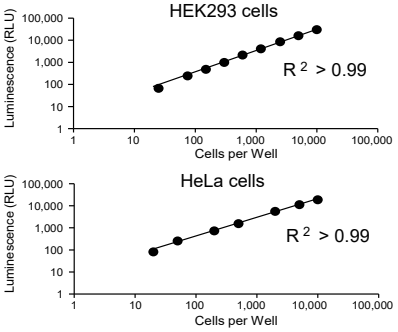
ATP-Luciferase Cell Viability Assay Solution (1.0mL×10) 1set [A3495]

Advantages

- **Premixed for quick and easy use. No need to remove medium. Results in only 10 minutes.**
- **High-sensitivity, linear (R^2 > 0.99) quantification of cell numbers over 4 orders of magnitude (20 – 10,000 cells per well of a 96-well plate).**
- **Red-to-orange color change upon application helps keep track of large numbers of samples.**

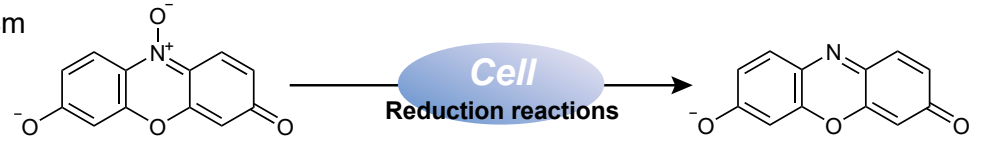
Directions for Use

1. Thaw assay reagent (A3495 or A3519) on ice.
2. Add volume of thawed assay reagent equivalent to volume of culture medium in each well.
3. Pipette gently to mix, incubate at room temperature for 10 minutes.
4. Measure chemiluminescence.



Resazurin (Ready-to-use solution) [for Cell proliferation assay] 25mL [R0195]

Mechanism

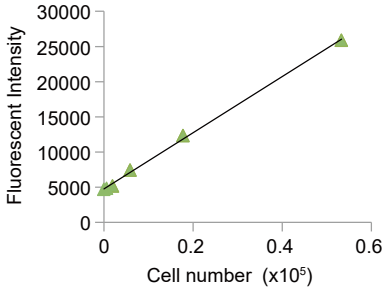


Ex: 540-570 nm, Em: 590 nm

Resazurin can be used to determine relative cell number, useful in the quantification of such phenomena as cell proliferation, viability, and cytotoxicity. When added to viable cells, blue, non-fluorescent resazurin undergoes a conversion to a highly fluorescent derivative. Both resazurin and its derivative are water soluble and have low toxicity, making it simple to handle and eliminating the washing, fixing, and extraction steps required for other commonly used cell proliferation assays.

Application: Cell viability assay by R0195

1. Add R0195 at a volume equal to 10% of the cell culture medium volume.
2. Return cells to the incubator and continue the incubation for 2-24 hours.
3. Measure the fluorescence intensity using 540-570 nm excitation and 590 nm emission wavelengths.
   *Absorbance measurement may also be used in place of fluorescence measurement to estimate cell number – use a 570 nm filter.



MTT [for Biochemical Research] 200mg / 1g [M3297]

MTT Solution [for Cell proliferation assay] (1mL×5) 1set [M3353]

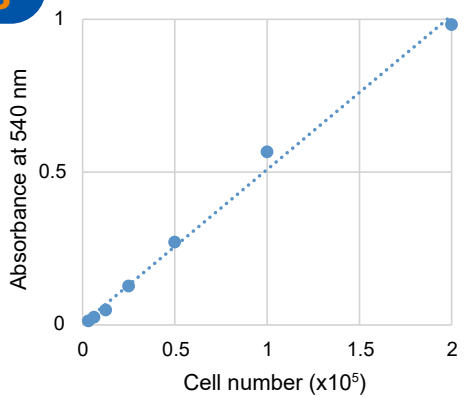
MTT is a type of tetrazolium salt that is converted into formazan when taken up by living cells. This formazan can be dissolved in DMSO and the absorbance at 540 nm can be used to estimate the relative number of cells. M3353 consists of 5.0 mg/mL MTT dissolved in PBS.

Cells stained by M3353

Application: Cell proliferation assay by M3353

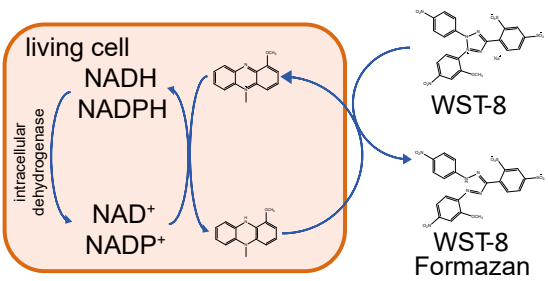
1. Seed 100 µL of cells in a 96-well plate, and Incubate overnight in an incubator.
2. Add 10 µL of MTT solution [M3353] to each well.
3. Incubate for 2-4 hours until a color change is seen.
4. Remove medium, taking care not to inhale the formazan.
5. Add 100 µL of DMSO and dissolve the formazan.
6. Measure the absorbance at 540 nm.

Please adjust staining time and volume as needed.
Preliminary tests should be performed.



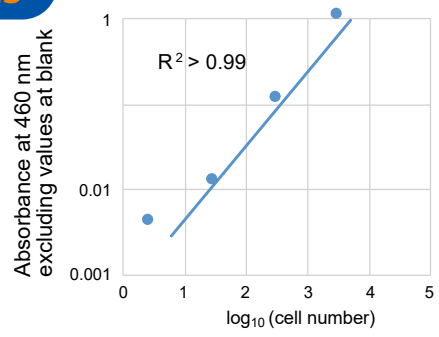
WST-8 Reagent [for Cell Proliferation Assay] 1mL [W0023]

W0023 is supplied as a ready-to-use reagent for the measurement of relative cell number. Reduction of the water-soluble tetrazolium salt WST-8 by the cell membrane-permeable electron carrier 1-MePMS converts it to a highly colored form which can be detected using a plate reader fitted with a 460 nm absorbance filter. As 1-MePMS is itself reduced by intracellular NADH/NADPH, and NADH/NADPH concentration is relatively constant over the course of the cell cycle, the amount of reduced WST-8 and therefore the degree of color change can be used to determine relative cell number.



Application: Cell proliferation assay using W0023

1. Seed 100 µL of cells in a 96-well plate, and incubate overnight in an appropriate incubator.
2. Add 10 µL of WST-8 Reagent [W0023] to each well.
3. Incubate for 1-4 hours until a color change is seen.
4. Measure the absorbance at 460 nm.



Mitochondrial Isolation Reagent

Mitochondrial Isolation Kit

1kit [M3527]

M3527 can be used to easily isolate mitochondria from cultured mammalian cells and tissue.
Isolated mitochondria can be used for downstream analyses such as Western blotting.

Kit Components

- **Mitochondrial Isolation Reagent A for approx. 30 samples**
- **Mitochondrial Isolation Reagent B for approx. 30 samples**
- **Mitochondrial Isolation Reagent C for approx. 30 samples**

Application: Isolation from Mouse-Derived Cells

1. Collect cells by centrifugation and remove the supernatant without disturbing the cell pellet. Use a cell scraper for detaching adherent cells.
2. Resuspend cells in Mitochondrial Isolation Reagent A and vortex for 5 seconds. Add 400 µL per 1 x 10^7 cells.
3. Incubate on ice for 2 minutes.
4. Add Mitochondrial Isolation Reagent B to the cell suspension and vortex for 5 seconds. Use 5 µL per 400 µL of suspension from step 2.
5. Incubate on ice for 5 minutes, vortexing once per minute.
6. Add Mitochondrial Isolation Reagent C and invert several times (do not vortex). Use a volume equal to the volume of Reagent A used in step 2.
7. Centrifuge at 700 x g for 10 minutes at 4°C.
8. Transfer the supernatant to a new tube and centrifuge at 12,000 x g for 15 minutes at 4°C.
9. Collect the supernatant (cytoplasmic fraction) and add 300 µL of Mitochondrial Isolation Reagent C to the pellet (mitochondria).
10. Centrifuge at 12,000 x g for 5 minutes at 4°C and remove the supernatant.
11. Use mitochondria (pellet) for downstream experiments.

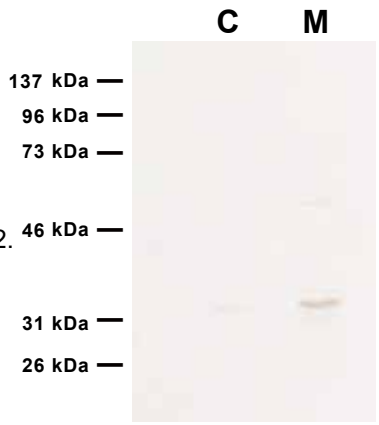


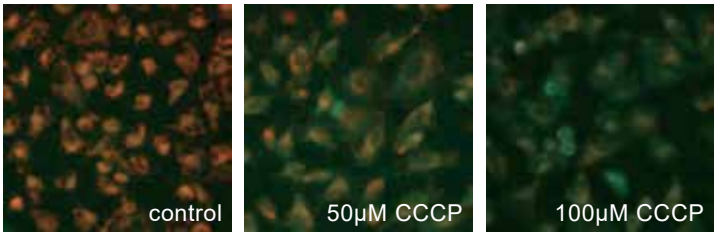
Figure.
Western blotting of M3527-isolated cytoplasmic fraction (C) and mitochondria (M), detected with VDAC1/2 antibody.
Mitochondrial proteins were extracted with RIPA Buffer.
M3527 allows for highly efficient purification of mitochondria

Mitochondrial Membrane Potential Assay Reagent

JC-1 Mitochondrial Membrane Potential Assay Kit

1kit [J0043]

J0043 (JC-1 Mitochondrial Membrane Potential Assay Kit) is used to provide a visual indicator of mitochondrial membrane potential. J0043 contains JC-1, whose monomeric green fluorescence converts to red fluorescence upon aggregation inside intact mitochondria, and CCCP, a mitochondrial depolarizing agent.



Fluorescence microscope image of HeLa cells stained with J0043

Nuclear / Cytoplasmic Fractionation Reagent

Nuclear / Cytoplasmic Fractionation Kit

1kit [N1208]

N1208 can be used for convenient fractionation of nuclear and cytoplasmic proteins from cultured mammalian cells. By using the three reagents included, quick collection of each fraction can be achieved. The extracted proteins can be used directly in downstream applications such as Western blotting.

Kit Components

- **1X Cytoplasmic Extraction Buffer**
  for approx. 50 samples
- **20X Detergent Solution**
  for approx. 50 samples
- **1X Nuclear Extraction Buffer**
  for approx. 50 samples

Application of Nuclear / Cytoplasmic Fractionation Kit [N1208]

[For Cytoplasmic Extraction]

1. Collect cells and wash cells twice with ice cold PBS. Use a cell scraper for adherent cells.
2. Resuspend the cells gently in cold 1X Cytoplasmic Extraction Buffer. Add 400 µL of the buffer to 100 µL of the cell volume (~ 1 x 10^7 cells).
3. Incubate on ice for 10 minutes.
4. Add 20X Detergent solution to the cell suspension and vortex vigorously for 10 seconds. Use 25 µL of 20X Detergent Solution per 500 µL of the solution from step 3.
5. Centrifuge at 800 x g for 10 minutes at 4 °C.
6. Transfer the supernatant (cytoplasmic fraction) to a new tube for downstream experiments.

[For Nuclear Extraction]

7. Resuspend the pellet in cold 1X Nuclear Extraction Buffer. Add 100 µL of 1X Nuclear Extraction Buffer per 100 µL pellet.
8. Incubate the pellet solution on ice for 20 minutes, vortexing every 5 minutes.
9. Centrifuge at 15,000 x g for 10 minutes at 4 °C.
10. Transfer the supernatant (nuclear fraction) to a new tube for downstream experiments.

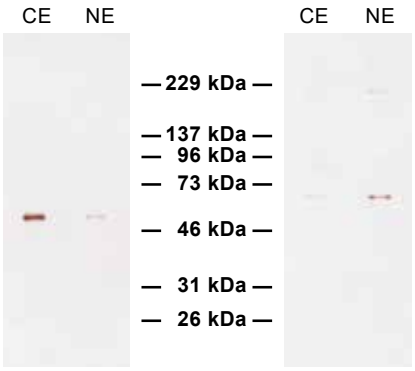


Figure.
Western blotting of cytoplasmic fraction (CE) and nuclear fraction (NE) extracted with **N1208**, detected using α-Tubulin antibody (left) and Lamin B1 antibody (right). Excellent separation was achieved.

Extraction Buffer for *E. coli* and Yeast

E.coli / Yeast Protein Extraction Buffer

100mL [Y0021]

This product is supplied as a ready-to-use solution to lyse *E. coli* / yeast cells. By suspending cells in this buffer [Y0021] and then centrifuging them, the supernatant containing proteins can be obtained. Extracted protein can be used in downstream applications such as Western blotting.

Application

1. Centrifuge *E. coli* at 5000 x g and yeast at 3000 x g for 10 minutes. Remove as much medium as possible from pellet.
2. Resuspend pellet in Y0021. Add 2-4 mL of Y0021 per gram pellet.
3. Mix at room temperature for 10 minutes.
4. Centrifuge *E. coli* at 15000 x g and yeast at 14000 x g for 10 minutes.
5. Collect the supernatant and measure the protein concentration.
6. Analyze them directly by Western blotting.

Protein Quantification

	When using Y0021	When using PBS
E. coli K-12	3.90 mg/mL	0 mg/mL
S. Cerevisiae Y-12632	2.79 mg/mL	0 mg/mL

Western Blotting

The extracts were transferred to a PVDF membrane after electrophoresis.
RecA in *E. coli* (left, 38 kDa) and Rad51 in *S. cerevisiae* (right, 43 kDa) could be extracted efficiently.

Extraction Buffer for Nervous Tissue

Nervous Tissue Protein Extraction Buffer

100mL [B6279]

This product is supplied as a ready-to-use solution to lyse nervous tissue. By suspending tissue in this buffer [B6279] and then centrifuging them, the supernatant containing proteins can be obtained. Extracted protein can be used in downstream applications such as Western blotting.

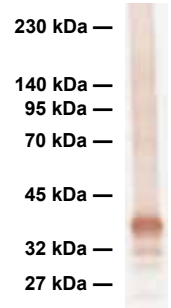
Application

1. Wash mouse brain twice with PBS.
2. Weigh samples. For each gram of sample, add 10 mL of B6279 and homogenize.
3. Incubate on ice for 10 minutes.
4. Centrifuge the sample at 10000 x g for 10 minutes at 4 °C.
5. Collect the supernatant and measure the protein concentration.
6. Analyze them directly by Western blotting.

Protein Quantification

When using B6279	When using PBS
3.63 mg/mL	1.59 mg/mL

Western Blotting



The extracts were transferred to a PVDF membrane after electrophoresis.
Synaptophysin (38 kDa) could be extracted from mouse brain efficiently.

Extraction Buffer for Mammalian Cells

RIPA Buffer (Ready-to-use) [for Protein extraction] 100mL [R0246]

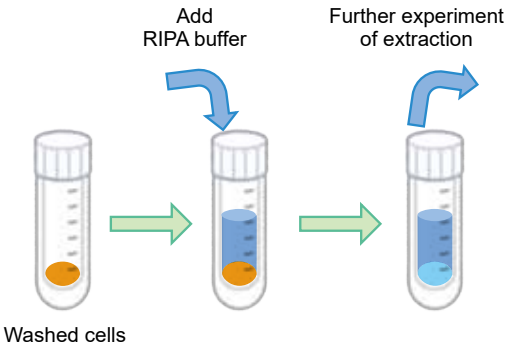
This product is supplied as a ready-to-use solution for the lysis of the cultured mammalian cells. Proteins can be extracted by adding this buffer [R0246] to the cells and the extract can be used directly for further analysis such as western blotting. This product does not include protease inhibitors. Please add a protease inhibitor cocktail, if necessary.

Application

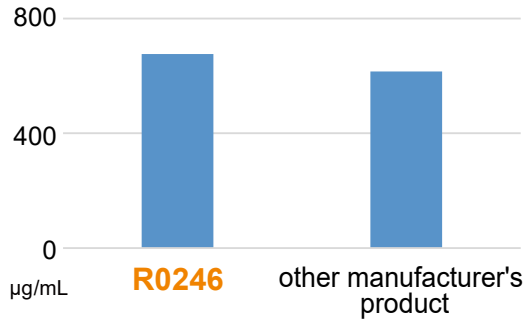
Add the following protease inhibitors to RIPA buffer [R0246].

Leupeptin	10 µg/mL
Pepstatin A	1 µg/mL
Aprotinin	3 µg/mL
AEBSF	1 mM

1. Wash the cultured mouse myeloma-derived cell sp2/0 twice with PBS.
2. Remove PBS and add 200 µL of either cold RIPA buffer [R0246] containing protease inhibitors or the other manufacturer's RIPA buffer containing the same protease inhibitors to 1.0 x 10^6 cells.
3. Incubate the cells for 15 minutes on ice.
4. Centrifuge the cells at 10000 x g for 10 minutes at 4 °C.
5. Measure the protein concentration of the supernatants.
6. Analyze the supernatants using western blotting.



Extracted Protein Concentration



The extracts were transferred to a PVDF membrane after electrophoresis.
Anti-β actin antibody was used for detection.
Equal or better detection was observed than that of the other manufacturer's product.

Related Products

100×Protease Inhibitor Cocktail (EDTA free) 1vial [P2949]
100×Protease Inhibitor Cocktail 1vial [P2976]

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